### **DISPOSAL OF USED DIP SLIDES**

Used Dip Slides can be incinerated, autoclaved or soaked overnight in a suitable disinfectant, such as hypochlorite or a phenolic or quaternary ammonium compound product.

### STORAGE OF DIP SLIDES

Dip slides can be stored at 10 -  $25^{\circ}$ C. Refrigeration at  $4^{\circ}$ C is acceptable but may result in the release of water condensation. Do not freeze. Do not use Dip Slides beyond the stated expiry date, or if the products show signs of deterioration.







# Dip Slide for Industrial Use

000 «Диаэм»

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### **OXOID DIP SLIDES**

The Oxoid Dip Slide is made of easily disposable plastic. Its raised edges ensure an even thickness of culture medium. It has a moulded grid, which makes colony counting easier, and is shaped to ensure drainage of fluid from the slide. The surface area of the agar layer on each side is 1000 sq.mm.

Dip Slides are simple to use both for the operator and where laboratory facilities are limited. They are ideal for the monitoring of oil and water emulsions, and are inexpensive for carrying out microbiological surveys.

### THE CAP

The bottle cap is an integral part of the Dip Slide assembly, forming a convenient handle by which it may be held without risk of touching the culture medium. Contamination with unwanted organisms that can increase the final count is therefore less likely.

### THE TECHNIQUE

The Oxoid Dip Slide is coated on both sides with a solid culture medium. Different media can be carried on each side. Both sides should be brought into contact with the surface or fluid to be investigated.

For fluids, the Dip Slide can either be dipped into the fluid or a little of the fluid can be poured over both sides of the slide. Be careful to wet the total surface of each side. Allow excess fluid to drain off for a few seconds and return the slide to the bottle. Do not incubate the test fluid with the dip slide in the bottle - a satisfactory count will **not** be achieved.

For checking the cleanliness of surfaces, each side of the coated slide should be pressed flat on the surface using enough pressure to bend the stem slightly. Return the slide to the bottle.

Incubate for a suitable time at the given temperature (as detailed in the table below) and count colonies. Use the grid pattern in the slide to sub divide the total area and make the counting easier. Oxoid Dip Slides are available for the following purposes:

Test	Code	Dip Slide Use
For the total count of aerobic bacteria	DS0147A	This carries TTC Red Spot Medium which is yellow in colour and transparent. Developing bacterial colonies alter the TTC and they appear as red spots. Incubate at 30°C - 37°C for 18-48 hours.

Test	Code	Dip Slide Use
For the total aerobic count and total yeast and mould count	DS0155A	This carries TTC Red Spot Medium on one side (see DS0147A) and Malt Extract Agar on the other side for yeast and mould isolation. Incubate at 25°C - 30°C for 2-5 days.
	DS0170A	On one side, the slide carries Plate Count Agar, for total aerobic counts. On the other, it carries OGYE Agar for the enumeration of yeasts and moulds. Incubate at 25°C - 30°C for 2-5 days.
For total and coliform bacterial count	DS0166A & DS0167A	These carry Plate Count Agar (PCA) on one side and MacConkey Agar No. 3 on the other. PCA will detect total aerobic counts, and MacConkey Agar No. 3 detects faecal coliforms, inhibiting gram positive cocci. The media of DS0167A also contain germicide inhibitors.  Incubate at 30°C - 37°C for 18-48 hours.
For total and Enterobacteriaceae bacterial count	DS0168A & DS0169A	These carry PCA on one side and Violet Red Bile Glucose Agar (VRBGA) on the other. PCA will detect total aerobic count and VRBGA will give purple red colonies for Enterobacteriaceae. The media of DS0169A also contain germicide inhibitors. Incubate at 30°C - 37°C for 18-48 hours.

### **EVALUATION OF RESULTS**

### Comparison of growth on Dip Slide

